

REVIEW ARTICLE

Pharmaceutics for the anaesthetist

R. D. MacPherson

Senior Lecturer, The Department of Anaesthesia and Pain Management, Royal North Shore Hospital, St Leonards, NSW 2065, Australia

Summary

Pharmaceutics is that branch of science concerned with the manufacture and formulation of pharmaceutical products, and is a subject almost exclusively in the domain of pharmacists and those concerned with pharmaceutical manufacture. However, there are some aspects of pharmaceutics that are of particular importance to the anaesthetist, such as the pharmacology of the various preservatives, antimicrobials and other additives found in anaesthetic products, and an understanding of basic processes such as emulsification and lyophilisation. This review aims to survey those areas.

Correspondence to: Dr R. D. MacPherson

E-mail: rmacpher@doh.health.nsw.gov.au

Accepted: 1 May 2001

There is little information in the anaesthetic literature on drug additives, and, indeed, it appears that only one previous significant summary has been published [1].

Anaesthetic products can contain a wide range of pharmaceutical additives that are included for a number of reasons. Principal among these is the need to prevent contamination of the product and to maintain potency by reducing its rate of breakdown, although they are also used to modify tonicity or pH, or to stabilise the product. The most common additives used in pharmaceutical products, which are discussed in this paper, are listed in Table 1.

Solvents

Water

Water is the most common and appropriate solvent used in drug preparation. It is tasteless, free from irritating qualities and is pharmacologically inert [2]. Despite these attributes, its use is not without a number of attendant problems. For example, drugs in solution are liable to breakdown, especially by hydrolysis, and the solutions are easily contaminated by micro-organisms.

Most drugs are water-soluble to some degree, depending on the relative polarities of solvent and solute, and this can be determined numerically by calculation of the dielectric constant, which is measured by placing a sample of the substance to be studied between two plates attached

to a voltage source and measuring the capacitance between the plates (C_x), which is then compared to the capacitance when a vacuum (C_0) occupies the space. The dielectric constant (ϵ) is determined by calculating C_x/C_0 . Substances with dielectric constants > 50 are generally classified as polar; those with constants within the range of 1–20 are non-polar; those with intermediate dielectric constants are referred to as semipolar (Table 2).

The high dielectric constant of water ($\epsilon = 78$) confirms it as a polar solvent, and it is this feature that facilitates the dissolution of many molecules, especially polar or charged species.

Non-aqueous solvents

Vehicles other than water are used for drug formulation for two reasons. The first is to solubilise drugs, which by virtue of the presence of non-polar or hydrophobic characteristics are relatively insoluble in aqueous vehicles. The second relates to product stability, since, generally, the use of non-aqueous vehicles with lower dielectric constants will reduce the rate of hydrolytic decomposition (see below). Most non-aqueous vehicles, such as glycerol, ethanol and members of the glycol family, are usually used in combination with water.

Propylene glycol

Glycols are dihydric alcohols derived from natural gas and are widely used in foodstuffs and in industrial products

Table 1 Principal pharmaceutical vehicles and additives.

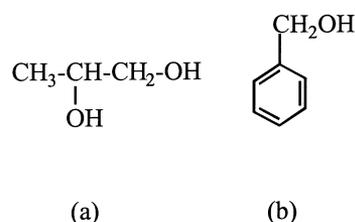
Vehicles	Water Propylene glycol Benzyl alcohol
Emulsions	
Surfactants	Polyoxyethylated castor oils Lecithin
Cyclodextrins	
Anti-oxidants	Alkyl-substituted phenols Sulphurous acid salts Edetic acid Lecithin
Preservatives	Benzalkonium and benzethonium chlorides Chlorbutol Parabens

such as antifreeze and brake fluid. As might be expected from their chemical structure (Fig. 1a), glycols are miscible with water in all concentrations. Ethylene glycol is too toxic for human use, being metabolised by ethanol dehydrogenase, eventually to form the toxic oxalic acid. However, propylene glycol (more correctly called propylene glycol monomethyl ether) is an agent of generally low toxicity [3, 4] that is used as a solubilising agent and for its mild preservative action. After administration, it is metabolised to lactic and pyruvic acids, and although uncommon, acidosis and serum hyperosmolality have occasionally been reported after high doses [5, 6]. Rapid intravenous administration of products containing propylene glycol has been associated with the development of hypotension and arrhythmias – a further reason why intravenous preparations of propylene glycol-containing agents, especially phenytoin and digoxin, should be administered slowly. Products using propylene glycol as a vehicle are also liable to cause significant thrombophlebitis, which in some studies has had an incidence as high as 60% [7], often with serious consequences [8]. Such findings have led manufacturers to reformulate propylene glycol-containing products such as diazepam into injections containing less irritant additives.

Preparations of the intravenous induction agent etomidate can contain up to 35% propylene glycol, and

Table 2 Dielectric constants (ϵ) of various solvents measured at 20 °C.

Compound	Dielectric constant
N-methylformamide	190
Water	78
Glycerine	46
Methanol	33
Propylene glycol	32
Castor oil	4.6
Olive oil	3.1

**Figure 1** Chemical structure of non-aqueous solvents: (a) propylene glycol, (b) benzyl alcohol.

it has been suggested that the solvent in some formulations may be responsible for pain on injection. Although the mechanism is unclear, it has been suggested that propylene glycol has a direct effect on the vasculature, resulting in local tissue and endothelial cell damage [9, 10], an effect that has been noted previously with other drugs [11].

Benzyl alcohol

In common with other drug additives having multiple actions, benzyl alcohol (Fig. 1b) has two applications in pharmaceutical formulation. In low concentrations of up to 2% it is used as a preservative and antimicrobial, and is employed in preparations of phenobarbitone, d-tubocurarine and some formulations of midazolam and diazepam. In concentrations of greater than 5%, it is used as a non-aqueous solvent.

Its toxicity is dose-related and was responsible for a number of deaths some years ago when benzyl alcohol-preserved saline was given to neonates [12–15]. Benzyl alcohol, which is metabolised to benzoic acid and thence conjugated with glycine to form hippuric acid, also has a number of cardiovascular effects. Animal experiments have confirmed that in sufficient doses its use will result in hypotension by both peripheral vasodilatation and myocardial depression [16–18].

Furthermore, it also possesses local anaesthetic properties – a fact that has been recognised for some time [19]. There has recently been a resurgence of interest in its use for this indication, with some promoting its use intradermally before intravenous cannulation, as it has been suggested that it has an efficacy similar to that of lidocaine but with less pain on injection [20, 21]. However, its use in this role could be limited because of its capacity to cause contact dermatitis and sensitisation [22–24], a response seen with a higher incidence in the population who also demonstrate aspirin allergy [25, 26].

Emulsions

An emulsion is a two-phase system, consisting usually of a pair of immiscible liquids, one of which is dispersed in the

form of small droplets throughout the other. As anybody who has made oil and vinegar salad dressing or mayonnaise will know, such systems are inherently unstable. The reason for this is not poor culinary skill but the laws of thermodynamics. When a dispersed phase is broken down to very small droplets, the result is an increase in total surface area of enormous proportions. For example, 1 ml of oil broken down to fine globules and then dispersed in water results in an interfacial surface in the order of some 500 m², which in turn creates a significant increase in the interfacial energy of the system [27].

In order to increase the stability of such an emulsion system, additives called 'emulsifying agents' are employed. This is a general term embracing a range of heterogeneous substances that stabilise emulsion systems by maintaining droplets in the dispersed phase, so preventing their coalescence. They include the surfactants and the hydrocolloids, and achieve their effects through a number of processes. Some form physical or chemical barriers around the dispersed droplets, or impart an electric charge to the external surface of the droplets so that they tend to repel one another. Others reduce the interfacial tension between the two phases, which in turn lowers the interfacial energy within the emulsion.

Emulsions are defined as being of the oil-in-water (o/w) type, where the oil is the droplet, also known as the internal or discontinuous phase, and is dispersed through an aqueous phase. The water-in-oil (w/o) type is where the water is in droplet form in an oily dispersion medium, also called the external or continuous phase. Which of these two forms of emulsion will be formed is primarily dependent on the relative mass of the phases and the nature of the emulsifying agent added. Of the commercial fat emulsions available, soya bean oil is the most common and is used in the emulsions of both propofol and diazepam, with purified egg yolk phospholipids as the emulsifier.

Despite the use of stabilising agents, emulsions remain unstable preparations, and are particularly sensitive to storage conditions, changes in temperature and added molecules. The aqueous phase is prone to contamination and usually contains a water-soluble preservative, while the oil phase is prone to rancidity, which necessitates the addition of a lipophilic anti-oxidant such as tocopherol.

Propofol

Propofol (Diprivan, AstraZeneca) is one of the most widely used emulsions in anaesthetic practise. It was originally marketed as an o/w emulsion using soya bean oil as the oil phase and egg lecithin as the emulsifying agent. While this has remained the standard formulation,

there have recently been some changes in other aspects of its manufacture which will be discussed elsewhere.

Propofol causes pain on intravenous injection, a phenomenon that has been extensively reviewed and investigated [28, 29]. The mechanism for this is clearly complex and may be multifactorial. While no firm consensus has been reached, some studies have suggested that the lipid solvent is responsible through activation of the plasma kallikrein system, which results in the production of bradykinin [30]. However, this has not been a consistent finding.

As a general rule, the addition of any other drug to an emulsion is contraindicated. There is a real risk that the admixture of other substances, especially charged species, can interact with either the lipid phase or the emulsifying agents, resulting in emulsion destabilisation [31]. Anaesthetists have taken to adding a wide range of drugs to propofol. These have included the local anaesthetics lidocaine, procaine and prilocaine, in an effort to decrease pain on injection [32–34], as well as thiopental [35]. While the addition of local anaesthetics has not been associated with any untoward effects to date, the stability of thiopental/propofol mixtures appears limited by emulsion stability [36].

Surfactants

Certain molecules possess structural features making them liable to accumulate at the boundaries between heterogeneous or immiscible phases and so reduce the interfacial tension between them. They are called surface-active agents, or surfactants, and permit the solubilisation of substances that would otherwise be impossible by acting as stabilising agents in emulsions.

Chemically they are amphiphiles, possessing both hydrophilic and lipophilic regions. The lipophilic portion is almost universally a long chain hydrocarbon, which may or may not contain an aromatic group, while the hydrophilic portion is more variable and may be anionic, cationic or non-ionic in nature. Surfactants tend to surround individual molecules of the internal phase and so form 'monomolecular' emulsions. Although there is a wide range of surfactants available, most are used in topical or oral products. However, there are some surfactants of anaesthetic interest. The cationic agents benzalkonium and benzethonium chlorides, which are better known as preservatives, and lecithin, an anti-oxidant with surfactant properties, are discussed elsewhere.

Polyoxylated castor oils

The reaction between rinoleic acid (castor oil), or its derivatives, with ethylene oxide results in the formation

of a range of chemicals with surfactant properties. These are called polyoxylated castor oils.

Cremophor EL (also known as Polyoxyl 35 Castor Oil) is the proprietary name of one of these products, and is composed of a number of different molecules that contain both lipophilic and hydrophilic components. The isolated lipophilic extract was once also used as a pharmaceutical solvent under the trade name Tensid, Micellophor or ORPE (oleum ricini polyethoxylate), and was used in the formulation of propanidid injection [1].

The lipophilic portion of Cremophor EL comprises a triglyceride stem to which are attached approximately 40 oxyethylene units. It has been used as a surfactant in pharmaceutical preparations since the 1970s and was incorporated into preparations such as propofol, phytonadione (vitamin K) and alphaxalone/alphadolone. However, serious anaphylactoid reactions associated with its use eventually led to the withdrawal of many compounds in which it was found, although propofol was later successfully reformulated in a soya bean oil emulsion.

A number of mechanisms have been proposed to account for these reactions [37], including Type I hypersensitivity reactions, complement activation [38] and direct histamine release from mast cells. Although no longer used in anaesthetic products, its efficacy in solubilising highly lipophilic compounds has led to its use in parenteral preparations of a range of substances including cyclosporin, doxorubicin, teniposide and paclitaxel.

Anaphylactic and hypersensitivity reactions continue to be reported after the administration of these Cremophor EL-containing products [38–40], as well a range of other adverse effects including pancreatitis [41] and central neurotoxicity [42], many of which seem to be dose-related [43, 44]. Experiments in which either rat aortic rings or rabbit artery segments were exposed to Cremophor EL resulted in persistent contractions through a direct action on vascular smooth muscle [45, 46]. The significance of this is unknown, but could imply that intra-arterial injection of Cremophor-containing products could be harmful. Further investigations have shown that a similar vasoconstrictor effect of the vehicle on the pulmonary vein can be demonstrated [47]. Some researchers have proposed that these toxic reactions may be due not so much to the Cremophor itself but rather to the residual ethylene oxide or polymeric by-products produced as a result of the manufacturing process [48]. Additionally, there have been other interesting effects observed after the administration of Cremophor-containing drugs. The first was an observation that Cremophor EL favourably modifies the pharmacokinetics of a number of antineoplastic agents, with a resulting increase in antitumour efficacy of the drugs [49]. Another

unusual effect of Cremophor is its capacity to modulate multidrug resistance (MDR). Certain cells have the capacity to produce high concentrations of proline-glycoprotein, a membrane protein that acts as a pump to extrude a range of antineoplastic drugs from within the cell [37]. Importantly, some antineoplastic agents have been shown to induce this effect, thus reducing their own efficacy. However, several surface-active agents, including Cremophor EL, used in preparations of, for example, daunorubicin and paclitaxel, have been shown to reverse MDR [50], acting more as a synergist than an inactive surfactant [51].

Lastly, in studies where the ratio of blood and plasma paclitaxel concentrations were measured, the ratio was lower in Cremophor-treated patients, which was thought to be due to an interaction between the two products, resulting in paclitaxel becoming trapped in the Cremophor EL matrices, reducing its capacity to be transported to other sites [52]. Other researchers have suggested that this phenomenon of molecular trapping could be exploited as a means to reduce the haemolytic properties of some drugs.

Cyclodextrins

Another novel approach to solubilising lipophilic drugs is the use of cyclodextrins. These are cyclic oligosaccharides derived from starch, containing 6–8 glucopyranose units. The three-dimensional structure of the resulting molecule has been likened to a segment of a hollow cone, with a hydrophilic exterior surface and lipophilic interior surface into which individual molecules can become incorporated. Such a configuration is particularly useful for the encapsulation of highly hydrophobic molecules, which can then be rendered soluble in aqueous solution. A number of studies of the administration of propofol complexed with 2-hydroxypropyl- β -cyclodextrin have been completed, and these have shown similar pharmacological and anaesthetic properties to the commercial products currently available [53]. Some of these propofol-cyclodextrin products have already been patented, and may be destined for clinical trials in the near future [54].

Drug decomposition and contamination

In compounding drug products, and especially parenteral products, the challenge is to make a stable and appropriate formulation of a drug in which, amongst other things, the active drug will be soluble and resistant to the two main problems likely to occur with time: decomposition and contamination.

Decomposition

The most common consequence of decomposition or degradation is the diminution of product potency because

of a reduction in the concentration of the active ingredient [55]. A more serious, but less frequent, outcome is the formation of decomposition products, which may themselves be harmful. Prevention of decomposition is important since, with rare exceptions such as the formation of the coloured adrenochrome as a decomposition product of epinephrine, there is generally no physical evidence that a product may have undergone considerable reduction in potency, and the user is therefore unaware that such a process has occurred. Drugs in solution may decompose by many pathways, but chief among these are hydrolysis, oxidation, isomerisation and photochemical decomposition [56].

Hydrolysis and its prevention

Hydrolysis is the chemical reaction of a compound with water and is the most common method of drug degradation. Hydrolytic cleavage is promoted by the presence of hydroxyl and hydrogen ions found in aqueous solutions, although other basic and acidic species can also be involved, and essentially any drug in aqueous solution will be liable to hydrolysis to some degree. Compounds containing lactam groups, or ester and amide bonds, are particularly liable to hydrolytic breakdown, and so drugs such as atropine, procaine, cocaine and physostigmine, as well as penicillin and cephalosporin antibiotics, are primary candidates for hydrolytic degradation.

There are a number of approaches to reduce the rate of hydrolysis, most of which rely on reducing the amount of drug in the aqueous phase [56]. Such strategies include:

- Determining by laboratory experiments at what pH the product is most stable and adjusting the pH of the product accordingly by the use of buffers.
- Alteration of the dielectric constant by the addition of non-aqueous solvents such as glycerine or propylene glycol.
- Addition of agents to form complexes to increase stability.
- Solubilisation by surfactants.
- Modification of side chain substituents.
- Lyophilisation.

Some of these processes deserve further comment. Modifying the pH of the formulation to enhance drug stability relies upon determining the ion having the greatest effect on the hydrolytic reaction [57]. In general, the hydroxyl ions have the stronger effect, and so the pH range of minimum decomposition is often found to be 3–4. Reducing the rate of hydrolysis by modification of molecular structure is clearly a process fraught with problems, as it could result in reduction of pharmacological activity. However, through careful alteration of non-critical areas of the molecule, it is possible to produce

modifications that retain the required activity, but with enhanced stability.

A further means to reduce hydrolysis is by lyophilisation or freeze-drying. This is employed in both the food and the pharmaceutical industries and is said to have been first used, at least in principle, by the Incas several centuries ago. They preserved food by storage atop high mountains, where a combination of very low temperature and low pressure existed. Modern freeze-drying of pharmaceutical products such as vecuronium relies on the removal of water from solution by sublimation and desorption. It is a complicated process requiring several steps that take place within a chamber, wherein the temperature is first reduced so that the solution of the drug is frozen. Then the pressure is lowered to a point at which it is less than the vapour pressure of ice at that temperature. When these conditions coexist, the ice is converted directly to water vapour by sublimation, leaving behind the lyophilised powder. Naturally, such products require reconstitution before use.

Oxidation and its prevention

The term oxidation was formerly used to describe the reaction of a substance with oxygen, although it is more properly defined as a reaction involving the loss of electrons. It is a common means of chemical breakdown, and compounds prone to degradation by this method include vitamins, phenolic compounds such as propofol, and catecholamines. Reduction in the rate of oxidation of drugs can be accomplished either by removing (to as greater extent as possible) available oxygen in the pharmaceutical preparation, or by the addition of antioxidants.

Removal of oxygen

Removing oxygen from drug products and replacing it with less reactive gases such as nitrogen would seem an attractive strategy to reduce the rate of oxidation, and indeed propofol injection uses nitrogen to fill ampoule headspace. However, it is a process that sounds deceptively simple. As one author succinctly said: 'it is easy to remove most of the oxygen from a container, but very difficult to remove it all!' [57]. This is an important consideration, since even trace amounts of oxygen can be enough to cause the initiation of the oxidation process, since many such reactions proceed by 'auto-oxidation'. This is a process initiated by the interaction of ultraviolet radiation with trace amounts of oxygen resulting in the formation of free radicals, charged molecules containing an unpaired valence electron in the outer shell that makes them both reactive and unstable. The reaction is then propagated by the reaction of these free radicals with other parts of the drug molecule, which in so doing

creates further free radicals, and so on. The process is catalysed by trace amounts of heavy metal ions and is promoted by increased temperatures.

Anti-oxidants

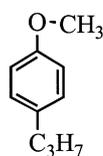
Anti-oxidants are generally classified into three groups [58]. The true anti-oxidants (also once appropriately called 'anti-oxygens') include alkyl-substituted phenols such as thymol, tocopherol, alkyl gallates and hydroxy anisole. They exert their anti-oxidant effect by interacting with free radicals, which makes them effective against auto-oxidation (above), but not against reversible oxidation–redox reactions. All of the substances are found in a range of parenteral products. Thymol, for example (Fig. 2a), is added to halothane to attenuate its rate of oxidation. Interestingly, propofol itself has been shown to have significant anti-oxidant properties [59], not an unexpected finding, as it shares the basic structure of a substituted phenol with other members of this group.

Reducing agents act as molecular sacrifices. Having a low redox potential, they are preferentially oxidised, thus leaving the active drug free from attack. They are also active against free radicals. Examples include ascorbic acid and the sulphurous acid salts, also known as sulphites.

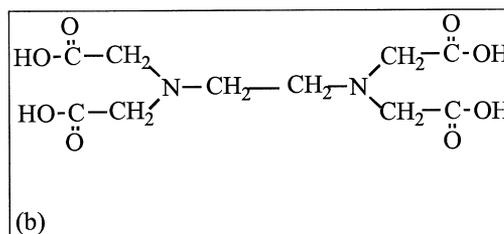
Sulphurous acid salts

Sulphurous acid salts or 'sulphites' comprise a range of agents including sulphur dioxide and sodium sulphite, bisulphite and metabisulphite, all of which release sulphur dioxide, the active moiety. Aminoglycosides and catecholamines are particularly liable to oxidation and often contain sulphiting agents, as do epinephrine-containing local anaesthetic solutions. In the last case, control of acidity is important, since at $\text{pH} > 4$ the two products react, forming the inactive epinephrine sulphonate [60].

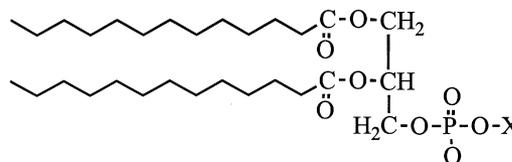
Sulphites are well known as mediators of hypersensitivity reactions [61], and the incidence of these reactions is dependent upon the route of administration [62]. Reactions are most likely after inhalation of sulphite-containing products [63–65], followed by oral ingestion. Such a reaction is usually characterised by wheeze, bronchospasm, pruritus and chest pain, and occurs almost exclusively in patients with pre-existing reactive airways disease and who are almost 10 times as likely to be steroid-dependent [66, 67]. The incidence of such reactions increases with age [68]. In the past, sulphites were common additives in inhaled bronchodilators and are now thought responsible for so-called 'paradoxical bronchospasm' sometimes seen after the use of inhaled sympathomimetics [63]. Reactions to parenteral injection of sulphites are rare but not unknown [69], and this response does not show the same predilection for steroid-dependent asthmatics [70]. In particular, total parenteral



(a)



(b)



with X as

Choline = Phosphatidylcholine

Ethanolamine = Phosphatidylethanolamine

Inositol = Phosphatidylinositol

Hydrogen = Phosphatidic acid

(c)

Figure 2 Chemical structure of the anti-oxidants (a) thymol, (b) edetic acid and (c) lecithin.

nutrition (TPN) solutions can contain large amounts of sulphites, with one report suggesting up to 950 mg of bisulphites per day could be administered [62], and that this should be a source of concern. Another suggested that the inadvertent administration of sulphite-containing TPN solutions over a number of days was responsible for the development of headache, vomiting, tachypnoea and coma in one patient [71]. Large doses in animal studies have also implicated sulphites in the development of renal cellular damage [72]. There has been a report of the development of generalised seizures in a patient given high doses of sodium bisulphate contained in intravenous morphine solution [73]. Dose-related toxicity is also theoretically possible with peritoneal dialysis fluids, some of which contain sodium metabisulphite in concentrations of 0.005–0.012% [74]. There has been a general desire by regulatory authorities to reduce the use of sulphites in pharmaceutical preparations [75, 76], and

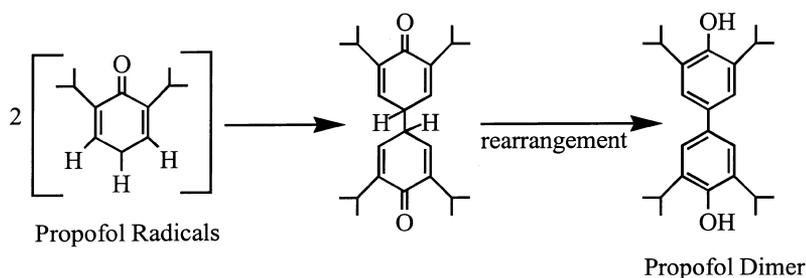


Figure 3 Proposed chemical transformation of propofol to its dimer, a process facilitated by the presence of metabisulphite (after [90]).

debate continues as to whether the sulphite concentration of some formulations of propofol could be potentially toxic [77, 78].

Anti-oxidant synergists enhance the activity of the 'true anti-oxidants', and are used in combination with them. This effect is mainly thought to come about through their interaction with heavy metal ions, which would otherwise catalyse the oxidation process. Examples of anti-oxidant synergists include lecithin, and citric, tartaric, and ethylenediaminetetra-acetic acid (EDTA, Edetic acid). Ethylenediaminetetra-acetic acid is a chelating agent used therapeutically in the treatment of heavy metal poisoning (Fig. 2b). It is used as an additive in some nebuliser solutions to chelate metal ions and to prevent discolouration of the solution [79]. It has also been used as an anti-oxidant synergist, but has insignificant antimicrobial activity. Ethylenediaminetetra-acetic acid has also been shown to have bronchoconstrictor properties [80, 81], and although the precise mechanism is unclear, it seems that the drug's calcium chelating properties may be responsible [82]. However, these effects have been seen only in concentrations far in excess of those used in human preparations and are unlikely to be clinically significant.

Lecithin (from the Greek *lekithos*, meaning 'egg yolk') is a phospholipid compound comprising a range of phosphatidyl esters such as phosphatidyl choline, ethanolamine and serine (Fig. 2c) together with varying amounts of other substances such as triglycerides and fatty acids [83]. It was originally obtained from eggs, although nowadays soya beans and other vegetables with a high lecithin content are also useful sources. Apart from its use as an anti-oxidant synergist, lecithin has important surfactant properties and is used as an emulsifying agent. It has also been proposed as an oral source of choline in the treatment of Alzheimer's disease and as an agent to reduce serum cholesterol, although in this case the promoters seem oblivious to the fact that lecithin is degraded in the gastrointestinal tract before having a chance to be absorbed.

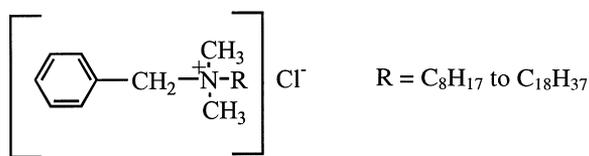
The question of whether propofol injection, because of its lecithin content, should be avoided in patients with egg

allergy is problematic. There have been no reports of adverse effects in the literature to date and although hypersensitivity reactions to both soya bean extract and lecithin have been demonstrated, they have only been reported when the allergens have been inhaled or ingested. Furthermore, patients who are designated as having a so-called 'egg allergy' generally demonstrate an IgE-mediated hypersensitivity to allergenic proteins found in egg whites. The derivation and chemical structure of egg lecithin would suggest that the risk of its administration to such individuals is very low.

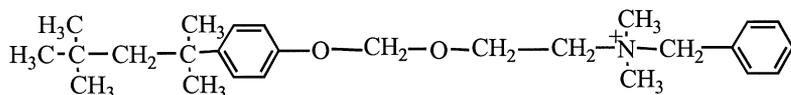
Recent re-formulations of propofol

Propofol was initially manufactured as a preservative-free product, and in some countries, such as Australia, this is still the case. However, as usage of the drug increased, reports of bacterial contamination of propofol emulsion also started to appear, with the suggestion that some cases of postoperative infection might have been due to the injection of contaminated product [84–86]. The concern that the preservative-free product could easily be contaminated through handling [87, 88] led the original patent holders, AstraZeneca, to reformulate the preparation. Antimicrobial studies demonstrated that EDTA at a concentration of 0.005% successfully retards microbial growth and has no adverse effects on the physicochemical stability of the product [89].

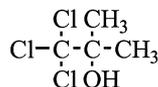
At about the same time that the EDTA preparation of propofol was being formulated, another manufacturer was marketing a sulphite-containing product. This product (propofol injectable emulsion 1%, Baxter Pharmaceutical Products) contains sodium metabisulphite 0.25 mg.ml^{-1} and is formulated in a slightly more acidic medium (pH 4.5–6.4) than the AstraZeneca product (pH 7.0–8.5). The release of these products has initiated a number of studies examining whether the new additives might have any effect on the stability of the emulsion product. A study by Baker [90] determined that both EDTA and sulphite-containing propofol products promoted the formation of a propofol dimer, formed after the propofol molecule had donated a hydrogen and resulting in the formation of a phenol-derived free radical species



(a)



(b)



(c)

(Fig. 3). While the propofol dimer was found in only trace amounts in the propofol–EDTA product (< 0.015%), its concentration in the sulphite–propofol product was significantly higher (0.18%). In addition, there have been reports of yellow discolouration in sulphite–propofol products, which the author suggests could be due to increased dimer production, which then undergoes further chemical transformation to coloured quinone species. Interestingly, both products are promoted by their respective manufacturers as being ‘preservative free’, since the concentration of both EDTA and sodium metabisulphite are below the United States Pharmacopoeia minimum standard that would otherwise require appropriate labelling.

Isomerisation

Isomerisation (or racemisation) refers to the process of conversion of a drug into its optical or geometric isomers [56]. Numerous studies on this topic have confirmed that potencies of drugs may differ substantially between isomers. As products begin being formulated as single isomers (such as ropivacaine), loss of potency through isomerisation must be considered. The rate at which racemic degradation occurs usually follows first-order kinetics and is dependent on temperature, solvent, catalyst and the presence of light [57].

Figure 4 Chemical structure of the preservatives (a) benzalkonium chloride, (b) benzethonium chloride and (c) chlorbutol.

Photochemical decomposition

Many compounds, especially those in solution, are light sensitive and will undergo complex multistep degradation after exposure to ultraviolet light. Fortunately, adequate protection can be afforded by storing products away from direct sunlight and by manufacturing them in dark amber glass. This effectively excludes ultraviolet radiation with wavelengths of less than 470 nm.

Preservatives

Preservatives are substances added to pharmaceutical products that prevent or inhibit the growth of microorganisms that may have been introduced during the manufacturing process. They are widely used in pharmaceutical products [91]. Almost all preservatives are weak acids with pKa’s in the order of 4–5. They will therefore

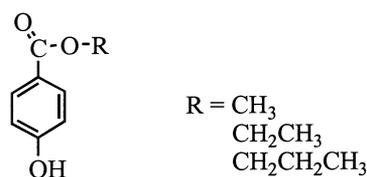


Figure 5 General chemical structure of the parabens.

be most effective in environments of an acid pH where, following the tenets of the Hendersen–Hasselbach equation, a greater proportion of the drug will exist in the un-ionised form and so be able to pass through microbial cell walls and membranes to exert an anti-bacterial effect. To achieve this end, acidifying agents are usually added to adjust the product pH accordingly.

Benzalkonium chloride (BAC)

Benzalkonium chloride, actually a mixture of quaternary benzyl-dimethylalkylammonium chlorides (Fig. 4a), is a common preservative found in eye and nose installations and in bronchodilator solutions [79]. Benzalkonium chloride causes reproducible bronchoconstriction that has a more rapid onset and prolonged duration compared with that caused by sulphites [92]. The mechanism may be direct histamine release, which occurs at concentrations $> 30 \mu\text{g}\cdot\text{ml}^{-1}$, almost the same as that suggested as the minimum effective dose as an antimicrobial [93]. In addition to BAC-induced bronchoconstriction, long-term sensitisation to BAC has also been reported. A chemically related preservative, benzethonium chloride (Fig. 4b), is used as a preservative in a number of formulations of ketamine injection. A recent study [94] has suggested that its inhibitory effect on muscarinic receptors could significantly contribute to the analgesic effect of ketamine.

Chlorbutol

Chlorbutol (chlorbutanol, Fig. 4c) has weak antibacterial and antifungal properties and has been used as a preservative, especially in ophthalmic preparations, and was formerly used in preparations of ketamine. A number of reports of hypersensitivity reactions have been received [95]. Chlorbutol also has weak sedative properties and there has been a case in which the chlorbutol added to morphine was thought to have contributed to the drug's sedative effects [96]. Other studies [97] have suggested that in doses of $10\text{--}100 \mu\text{g}\cdot\text{ml}^{-1}$, chlorbutol could activate the sodium–potassium pump in muscle preparations.

Parabens

The parabens are aliphatic esters of parahydroxybenzoic acid, from whence their name derives, and comprise methyl, propyl and butyl parabens (Fig. 5). Sodium benzoate and benzoic acid, although not parabens by strict chemical definition, are closely related structurally and possess a degree of cross-reactivity with members of the paraben class [98]. The parabens are preservatives that prevent the growth of fungi and yeasts, having less activity against bacteria. Some studies [99, 100] have suggested that parabens in general are weakly oestrogenic, although

the authors suggest that significant clinical problems are only likely to occur after long-term use of topically applied paraben-containing cosmetic products, and are unlikely to be of any concern after the systemic administration of the small amounts found in pharmaceutical products. Chronic use can also result in a reduction of cyclic adenosine monophosphate (cyclic AMP) through phosphodiesterase stimulation [101]. Reports of allergic reactions to parabens are uncommon, with one study estimating an incidence of approximately 0.3% [102]. However, another report [103] demonstrated that the incidence of intradermal reaction to prilocaine was reduced from 17% to 4% when paraben-free prilocaine was used, suggesting a slightly higher incidence of reaction.

Whether parabens have any direct vascular effects is unclear. Early studies suggested that parabens might have direct vasodilator activity on isolated human pial arteries and might be responsible for vasoactive effects of preparations of suxamethonium or naloxone [104, 105]. However, further studies failed to demonstrate any particular vasodilator properties of the group [106, 107].

Preservatives are biologically reactive substances, and their allergenic potential has been recognised for some time. One study that examined data from over 28 000 patch tests [108] concluded that the incidence of reactions to preservatives was: benzalkonium chloride 2%; benzyl alcohol 0.4%; sodium benzoate 0.4%; chlorbutol 0.1%; parabens 1.6%. In any case, their low allergenic potential and relatively high efficacy have made parabens the most popular preservatives in use today [109].

Adjustment of tonicity and pH

Adjustment of the osmolality of parenteral products to achieve isotonicity is important to decrease pain on injection, the incidence of thrombophlebitis and possible haemolysis. Many of the additives already discussed make a considerable contribution to the osmolality of pharmaceutical preparations, thus rendering them hypertonic. This is not usually of any clinical significance unless the osmolality exceeds $550 \text{ mosm}\cdot\text{l}^{-1}$. Where solutions are hypotonic, solutions of sodium chloride of varying strengths are added to adjust tonicity, although other agents such as glycerol are sometimes used. The amount required is either calculated using appropriate tables or is determined directly by measurement of freezing point depression. Glycerol is a common additive used to increase osmolality in hypotonic products and is used in propofol injection for that purpose.

As outlined in previous sections, adjustment of the pH of a drug product is performed for a wide range of reasons. It might be done for the purpose of promoting

drug solubility or stability, or to maintain the pH of a product in the acidic range where other additives, such as preservatives, are maximally active. Where pH adjustment is desired it is usual to use buffer pairs.

Adverse effects due to drug additives

The vast majority of additives used in pharmaceutical preparations are also widely used in the food industry. It is therefore far more likely that patients will be exposed to a wider variety, and to greater amounts, of these products as a result of ingestion of foodstuffs compared to the small amounts contained in pharmaceutical products [110, 111].

Adverse reactions can be dose-related or of an allergic or hypersensitive nature, in which case only a small amount will be needed to elicit a response in a susceptible individual. Although dose-related adverse responses to pharmaceutical additives are uncommon, there are at least four subgroups of patients who are at risk. These are: children, especially neonates; patients receiving TPN solutions; patients who for any reason are receiving long-term parenteral drug treatment, such as patients in intensive therapy units; those with chronic pain states who are using indwelling pump systems.

Intrathecal and epidural toxicity

Advances in the fields of chemotherapy and chronic pain management have resulted in an increase in the number of drugs introduced into the intrathecal space. Despite the fact that early work by Sjöberg [112] suggested the use of long-term intrathecal infusions containing preservatives was not associated with any histopathological damage, there have been continuing concerns about the relative safety of using non-preservative-free products [113, 114]. While there is evidence to suggest that the use of chlorbutol [115] or sodium bisulphite [116] under certain conditions can be linked to neurotoxic sequelae after intrathecal administration, such conclusions cannot be extrapolated to include all preservatives. For example, there have been a number of studies of the safety of paraben-containing solutions of neostigmine [20, 113, 117], the conclusions of which suggest that their administration is not associated with any obvious sequelae [117, 121].

Sulphites received attention in the 1980s when their accidental intrathecal administration in combination with the local anaesthetic 2-chlorprocaine resulted in a number of cases of permanent neurological impairment [116]. Investigations by a number of workers [122, 123] determined that it was the sodium bisulphite preservative which was responsible for the observed nerve damage, perhaps enhanced by the acidic pH of the solution [124].

There is also conflicting evidence concerning the safety of solutions containing benzyl alcohol given by the intrathecal route [125, 126], since under certain conditions their administration has been associated with neurotoxic sequelae [127, 128]. There have been insufficient studies performed to completely clear the drug for intrathecal use, and at least one group has suggested that more controlled studies should be conducted on aspects of its use near the spinal cord [114].

Similarly, it has also been advocated that any drug administered into the epidural space should also be preservative-free [129, 130]. Despite the general feeling that all drugs administered intrathecally or epidurally should be preservative-free [114], this is not always practically possible, as manufacturers are sometimes unable or reluctant to produce preservative-free products. Furthermore, the costs involved in the preparation of these products are often prohibitive, costing more than 10 times the cost of its preservative-containing counterpart [131]. Practitioners are therefore often faced with a choice between using preservative-containing products originally produced for systemic use, or not giving the drug at all. However, to complicate the picture further, reports of epidural inflammation following the implantation of long-term epidural catheters continue to occur despite the use of preservative-free solutions [132, 133], causing some to question the relative importance of demanding the use of preservative-free solutions when they are introduced into the epidural space [131].

Propylene glycol is a common additive in depot steroid injections such as methylprednisolone and triamcinalone, in concentrations of up to 3%. These products were never intended for spinal use [134] but have been introduced into the epidural space for decades for the treatment of a variety of conditions [135, 136]. A number of writers have suggested that pharmaceutical additives such as propylene glycol or benzyl alcohol might be responsible for cases of arachnoiditis occasionally seen after epidural steroid injection [137–139], although, as pointed out by Spaccarelli in an extensive review [140], most of these cases had complicating factors, making it difficult to draw any clear conclusions. Furthermore, a wide range of animal studies failed to demonstrate that either propylene glycol or benzyl alcohol had any direct neurotoxic effects [141–144].

Conclusions

The various additives used in pharmaceutical manufacturing represent a diverse range of chemical compounds with varied actions. Indeed, many drug additives exhibit more than one action. Importantly, information detailing the exact pharmaceutical excipients that are included in any

particular product can be difficult to obtain [145]. Such data are often omitted on drug labels, and may only be found by a careful examination of either the package insert or the manufacturer's prescribing information [146].

As long as 10 years ago, those involved in pharmaceutical manufacturing made the point that many drug additives were less than ideal and had a number of deficiencies which should be addressed, such as a high degree of sensitisation or dose-related toxicities [147, 148]. They concluded that new classes of preservatives were urgently needed. Unfortunately, there have been few advances in the development of new preservative agents in particular, or drug additives in general, in recent years, although the cyclodextrins offer an improved method of enhancing the solubility of highly lipophilic compounds which are found so often in anaesthesia.

Commercial pharmaceutical preparations usually contain at least one of the so-called 'additives' in combination with the active drug. Under normal circumstances, these additives have little pharmacological activity. Nevertheless, the anaesthetist should be aware of the rationale behind their inclusion, and of the circumstances under which they may exhibit toxicities.

Acknowledgments

I thank Ms Susan Ulstrup, who assisted with the preparation of the references for this manuscript, and Ms Bronwyn Fryirs, who computer-generated the structural formulae.

References

- Walsh R. Drug Preservatives and Solvents. In: Prys-Roberts C, Hug CC, eds. *Pharmacokinetics of Anaesthesia*. London: Blackwell Scientific, 1984: 321–45.
- Pernarowski M. Solutions, Emulsions and Suspensions. In: Gennaro AR, Gibson MR, Harvey SC, et al, eds. *Remington's Pharmaceutical Sciences*. Pennsylvania: Mack Publishing Co., 1975: 1436–53.
- LaKind JS, McKenna EA, Hubner RP, Tardiff R.G. A review of the comparative mammalian toxicity of ethylene glycol and propylene glycol. *Critical Reviews in Toxicology* 1999; **29**: 331–65.
- Cavender FL, Sowinski EJ, Glycols. *Patty's Industrial Hygiene and Toxicology*, Vol. 2F: Toxicology. Clayton GD, Clayton FE, eds, 4th edn. New York: Wiley, 1994: 4645–719.
- Cate JC, Hedrick R. Propylene glycol intoxication and lactic acidosis. *New England Journal of Medicine* 1980; **303**: 1237.
- York RC, Coleridge ST. Cardiopulmonary arrest following intravenous phenytoin loading. *American Journal of Emergency Medicine* 1988; **6**: 255–9.
- Mattila MAK, Ruoppi M, Korhonen M, Larni HM, Valtonen L, Heikkinen H. Prevention of diazepam-induced thrombophlebitis with Cremophor as a solvent. *British Journal of Anaesthesiology* 1979; **51**: 891–4.
- Schneider S, Mace JW. Loss of limb following intravenous diazepam. *Pediatrics* 1974; **53**: 112.
- Ruo W, Shay J, Attele A, Doenicke AW, Moss J. Propylene glycol damages vascular smooth muscle and endothelium. *Anesthesiology* 1992; **77S**.
- Doenicke AW, Roizen MF, Hoernecke R, Lorenz W, Ostwald P. Solvent for etomidate may cause pain and adverse effects. *British Journal of Anaesthesia* 1999; **83**: 464–6.
- MacPherson RD, McLeod LJ, Grove AJ. Intra-arterial thiopentone is directly toxic to vascular endothelium. *British Journal of Anaesthesia* 1991; **67**: 546–52.
- Anderson CW, Ng KJ, Andresen B, Cordera L. Benzyl alcohol poisoning in a premature newborn infant. *American Journal of Obstetrics and Gynaecology* 1984; **148**: 344–6.
- Menon PA, Thach BT, Smith CH, et al. Benzyl alcohol toxicity in a neonatal intensive care unit: incidence, symptomatology and mortality. *American Journal of Perinatology* 1984; **1**: 288–92.
- Hiller JL, Benda GI, Rahatzad M, Allen JR, Culver CV, Reynolds JW. Benzyl alcohol toxicity: impact on mortality and intraventricular hemorrhage among very low birth weight infants. *Pediatrics* 1986; **77**: 500–6.
- Benda GI, Hiller JL, Reynolds JW. Benzyl alcohol toxicity: impact on neurologic handicaps among surviving very low birth weight infants. *Pediatrics* 1986; **77**: 507–12.
- Dowdy EG, Holland WC, Yamanaka I, Kaya K. Cardioactive properties of d-tubo-curarine with and without preservatives. *Anesthesiology* 1971; **34**: 256–61.
- Reitan JA, James OF, Martucci RW. Cardiovascular effects of atropine sulfate preparations in vagotomized dogs under halothane anesthesia. *Anesthesia and Analgesia* 1977; **56**: 338–43.
- Yasaka WJ, Eichbaum FW, Oga S. Antiarrhythmic effects of solvents: III. Effects of propylene glycol and benzyl alcohol on contractile force isolated rabbit heart. *Cardiovascular Research* 1979; **13**: 717–22.
- Macht DIA. pharmacological and therapeutic study of benzyl alcohol as a local anesthetic. *Journal of Experimental Therapies* 1918; **11**: 263–79.
- Wilson L, Martin S. Benzyl alcohol as an alternative local anesthetic. *Annals of Emergency Medicine* 1999; **33**: 495–9.
- Fein JA, Boardman CR, Stevenson S, Selbst SM. Saline with benzyl alcohol as intradermal anesthesia for intravenous line placement in children. *Pediatric Emergency Care* 1998; **14**: 119–22.
- Verecken P, Birringer C, Knitelius A-C, Herbaut D, Germaz M-A. Sensitization to benzyl alcohol: a possible cause of 'corticosteroid allergy'. *Contact Dermatitis* 1998; **38**: 106.

- 23 Shunes E. Allergic dermatitis to benzyl alcohol in an injectable solution. *Archives of Dermatitis* 1985; **120**: 1200–1.
- 24 Kimura ET, Darby TD, Krause RA, Brodyk HD. Parenteral toxicity studies with benzyl alcohol. *Toxicology and Applied Pharmacology* 1971; **18**: 60–8.
- 25 Settupane GA. Adverse reactions to aspirin and related drugs. *Archives of Internal Medicine* 1981; **141**: 328–32.
- 26 Settupane GA. Aspirin and allergic diseases: a review. *American Journal of Medicine* 1983; **74**: 102–9.
- 27 Martin AN, Swarbrick J, Cammarata A. *Physical Pharmacy*. London: Henry Kimpton Publishers, 1973.
- 28 Picard P, Tramer MR. Prevention of pain on injection with propofol: a quantitative systematic review. *Anesthesia and Analgesia* 2000; **90**: 963–9.
- 29 Tan CH, Onsiong MK. Pain on injection of propofol. *Anaesthesia* 1997; **53**: 468–76.
- 30 Nakane M, Iwama H. A potential mechanism of propofol-induced pain on injection based on studies using nafamostat mesilate. *British Journal of Anaesthesia* 1999; **83**: 397–404.
- 31 Whately TL, Steele G, Urwin J, Smail GA. Particle size stability of intralipid and mixed total parenteral nutrition mixtures. *Journal of Clinical and Hospital Pharmacy* 1984; **9**: 113–26.
- 32 Scott RPF, Saunders DA, Norman J. Propofol clinical strategies for preventing pain on injection. *Anaesthesia* 1988; **43**: 492–4.
- 33 Nicol ME, Moriarty J, Edwards J, Robbie DS, A'Hern RP. Modification of pain on injection of propofol – a comparison between lignocaine and procaine. *Anaesthesia* 1991; **46**: 67–9.
- 34 Eriksson M. Prilocaine reduces injection pain caused by propofol. *Acta Anaesthesiologica Scandinavica* 1995; **39**: 210–13.
- 35 Prankerd RJ, Jones RD. Physicochemical compatibility of propofol with thiopental sodium. *American Journal of Health-System Pharmacy* 1996; **53**: 2606–10.
- 36 Chernin EL, Stewart JT, Smiler B. Stability of thiopental sodium and propofol in polypropylene syringes at 23 and 4°C. *American Journal of Health-System Pharmacy* 1996; **53**: 1576–9.
- 37 Dorr RT. Pharmacology and toxicology of cremophor EL diluent. *Annals of Pharmacotherapy* 1994; **28**: S11–4.
- 38 Szebeni J, Muggia FM, Alving CR. Complement activation by Cremophor EL as a possible contributor to hypersensitivity to paclitaxel: an in vitro study. *Journal of the National Cancer Institute* 1998; **90**: 300–6.
- 39 Kuiper RAJ, Malingre MM, Beijnen JH, Schellens JHM. Cyclosporine-induced anaphylaxis. *Annals of Pharmacotherapy* 2000; **34**: 858–61.
- 40 Millward MJ, Webster LK, Rischin D, *et al.* Phase I trial of cremophor EL with bolus doxorubicin. *Clinical Cancer Research* 1998; **4**: 2321–9.
- 41 Mills KM, Johnson DM, Middlebrooks M, Burton GV. Possible drug-associated pancreatitis after paclitaxel-cremophor administration. *Pharmacotherapy* 2000; **20**: 95–7.
- 42 Mross K, Hollander N, Hauns B, Schumacher M, Maier-Lenz H. The pharmacokinetics of a 1-h paclitaxel infusion. *Cancer Chemotherapy and Pharmacology* 2000; **45**: 463–70.
- 43 Badary OA, Al-Shabanah OA, Al-Gharably NM, Elmazar MM. Effect of Cremophor EL on the pharmacokinetics, antitumor activity and toxicity of doxorubicin in mice. *Anti-Cancer Drugs* 1998; **9**: 809–15.
- 44 Liao-Chu M, Theis JG, Koren G. Mechanism of anaphylactoid reactions: improper preparation of high-dose intravenous cyclosporine leads to bolus infusion of Cremophor EL and cyclosporine. *Annals of Pharmacotherapy* 1997; **31**: 1287–91.
- 45 Yaris E, Tuncer M, Kayaalp SO, Ilhan M. Direct vascular smooth muscle contractile effects of cyclosporin. A and its vehicle in rabbit isolated arteries. *Archives Internationales de Pharmacodynamie et de Therapie* 1994; **327**: 166–74.
- 46 Zengil H, Hodoglugil U, Guney Z. Effects of polysorbates and cremophor EL on vascular responses in rat aorta. *Experientia* 1995; **51**: 1055–9.
- 47 Mathieu P, Demers P, Elkouri S, Cartier R. The vasoconstrictor effect of cyclosporine on the pulmonary vein is mediated by its vehicle: the cremophor. (French). *Annales de Chirurgie* 1998; **52**: 821–6.
- 48 Windebank AJ, Blexrud MD, de Groen PC. Potential neurotoxicity of the solvent vehicle for cyclosporine. *Journal of Pharmacology and Experimental Therapeutics* 1994; **268**: 1051–6.
- 49 Badary OA, Abdel-Naim AB, Khalifa AE, Hamada FM. Differential alteration of cisplatin cytotoxicity and myelotoxicity by the paclitaxel vehicle cremophor EL. *Naunyn-Schmiedeberg's Archives of Pharmacology* 2000; **361**: 339–44.
- 50 Buckingham LE, Balasubramanian M, Emanuele RM, Clodfelter KE, Coon JS. Comparison of solutol HS 15, Cremophor EL and novel ethoxylated fatty acid surfactants as multidrug resistance modification agents. *International Journal of Cancer* 1995; **62**: 436–42.
- 51 Sparreboom A, van Tellingen O, Nooijen WJ, Beijnen JH. Nonlinear pharmacokinetics of paclitaxel in mice results from the pharmaceutical vehicle Cremophor EL. *Cancer Research* 1996; **56**: 2112–15.
- 52 Sparreboom A, van Zuylem L, Brouwer E, *et al.* Cremophor EL-mediated alteration of paclitaxel distribution in human blood: clinical pharmacokinetic implications. *Cancer Research* 1999; **59**: 1454–7.
- 53 Bielen S, Lysko G, Gough W. The effect of cyclodextrin vehicle on the cardiovascular profile of propofol in rats. *Anesthesia and Analgesia* 1996; **82**: 920–4.
- 54 Trapani G, Altomare C, Sanna E, Biggio G, Liso G. Propofol in anesthesia, mechanism of action, structure-activity relationships and drug delivery. *Current Medicinal Chemistry* 2000; **7**: 249–71.
- 55 Vermeire A, Remon JP. Stability and compatibility of morphine. *International Journal of Pharmaceutics* 1999; **187**: 17–51.
- 56 Florence AT, Attwood D. *Physicochemical Principles of Pharmacy*. London: Macmillan Press Ltd, 1988.

- 57 Lintner CJ. Stability of pharmaceutical products. In: Gennaro AR, Gibson MR, Harvey SC, et al, eds. *Remington's Pharmaceutical Sciences*. Pennsylvania: Mack Publishing Co., 1975: 1419–24.
- 58 Reynolds JEF, ed. *Martindale-the Extra Pharmacopoeia*. London: The Pharmaceutical Press, 1989.
- 59 Murphy PG, Myers DS, Davies MJ, Webster NR, Jones JG. The antioxidant potential of propofol (2,6-diisopropylphenol). *British Journal of Anaesthesia* 1992; **68**: 613–8.
- 60 Cartwright PD, Fyhr P. The manufacture and storage of local anaesthetics. *Regional Anaesthesia* 1988; **13**: 1–12.
- 61 Asmus MJ, Sherman J, Hendeles L. Bronchoconstrictor additives in bronchodilator solutions. *Journal of Allergy and Clinical Immunology* 1999; **104**: S53–60.
- 62 Golightly LK, Smolinske SS, Bennett ML, Sutherland EW, III, Rumack BH. Pharmaceutical Excipients Adverse Effects Associated with Inactive Ingredients in Drug Products (Part I). *Medical Toxicology* 1988; **3**: 129–65.
- 63 Goldfarb G, Simon R. Provocation of sulfite sensitive asthma. *Journal of Allergy and Clinical Immunology* 1984; **73**: 135.
- 64 Schwartz HJ. Sensitivity to ingested metabisulfite: variations in clinical presentation. *Journal of Allergy and Clinical Immunology* 1983; **71**: 487–9.
- 65 Prenner BM. Anaphylaxis after ingestion of sodium bisulfite. *Annals of Allergy* 1976; **37**: 180–2.
- 66 Schwartz JJ, Sher TH. Metabisulfite sensitivity in a patient without hyperactive airways disease. *Immunology and Allergy Practice* 1986; **8**: 308–11.
- 67 Bush RK, Taylor SL, Holden K, Nordlee JA, Busse WW. Prevalence of sensitivity to sulfiting agents in asthmatic patients. *American Journal of Medicine* 1986; **81**: 818–20.
- 68 Vandebossche LE, Hop WC, de Jonste JC. Bronchial responsiveness to inhaled metabisulfite in asthmatic children increases with age. *Pediatric Pulmonology* 1993; **16**: 236–42.
- 69 Baker GJ, Collett P, Allen DH. Bronchospasm induced by metabisulfite-containing foods and drugs. *Medical Journal of Australia* 1981; **2**: 614–16.
- 70 Smolinske SC. Review of parenteral sulfite reactions. *Journal of Toxicology-Clinical Toxicology* 1992; **30**: 597–606.
- 71 Abumrad NN, Schneider AJ, Steel D, Rogers LS. Amino acid intolerance during prolonged total parenteral nutrition reversed by molybdate therapy. *American Journal of Clinical Nutrition* 1981; **34**: 2551–9.
- 72 Akanji MA, Olagoke OA, Oloyede OB. Effect of chronic consumption of metabisulfite on the integrity of the rat kidney cellular system. *Toxicology* 1993; **81**: 173–9.
- 73 Meisel SB, Welford PK. Seizures associated with high-dose intravenous morphine containing sodium bisulfite preservative. *Annals of Pharmacotherapy* 1992; **26**: 1515–17.
- 74 Halaby SF, Mattocks AM. Absorption of sodium bisulfite from peritoneal dialysis solutions. *Journal of Pharmaceutical Sciences* 1965; **54**: 52–5.
- 75 Anonymous. *Sulfiting Agents: Labelling in Drugs for Human Use*. London: Department of Health and Human Services, Food and Drug Administration, 1986.
- 76 Challen RG. Sulphite content of Australian pharmaceutical products. *Medical Journal of Australia* 1990; **152**: 196–8.
- 77 Langevin PB. Propofol containing sulfite-potential for injury. *Chest* 1999; **116**: 1140–1.
- 78 Lakamp JE, Dobesh PP. Propofol and too much sulfite. *Chest* 2000; **118**: 277.
- 79 Beasley R, Fishwick D, Miles JF, Hendeles L. Preservatives in Nebulizer Solutions: Risks without benefit. *Pharmacotherapy* 1998; **18**: 130–9.
- 80 Downes H, Hirshman CA. Importance of calcium in citric acid induced airway constriction. *Journal of Applied Physiology* 1983; **55**: 1496–500.
- 81 Downes H, Hirshman CA. Calcium chelators increase airway responsiveness. *Journal of Applied Physiology* 1985; **59**: 92–5.
- 82 Hirshman CA, Peters J, Butler J, Hanifin HM, Downes H. Role of mediators in allergic and non-allergic asthma in dogs with hyper-reactive airways. *Journal of Applied Physiology* 1983; **54**: 1108–14.
- 83 Bowman WC, Rand MJ. *Textbook of Pharmacology*, 2nd edn. Oxford: Blackwell Scientific, 1980.
- 84 Bennett SN, McNeil MM, Bland LA, et al. Postoperative infections traced to contamination of an intravenous anesthetic, propofol. *New England Journal of Medicine* 1995; **333**: 147–54.
- 85 Nichols RL, Smith JW. Bacterial contamination of an anesthetic agent (editorial). *New England Journal of Medicine* 1995; **333**: 184–5.
- 86 Verber B, Gachot B, Bedos JP, Wolff M. Severe sepsis after intravenous injection of contaminated propofol (Letter). *Anesthesiology* 1994; **80**: 712–13.
- 87 Soong WAL. Bacterial contamination of propofol in the operating theatre. *Anaesthesia and Intensive Care* 1999; **27**: 493–6.
- 88 Sklar G. Propofol and postoperative infections. *Annals of Pharmacotherapy* 1997; **31**: 1521–3.
- 89 Boby DG, Russell AD, Jones CB. Selection of disodium edetate as the optimal antimicrobial additive for use in propofol emulsion. *Pharmacy and Therapeutics* 2000; **25**: 589–603.
- 90 Baker MT. Comparison of emulsion chemistry between sulfite-containing propofol with disodium edetate. *American Journal of Anesthesiology* 2000; **27** (6S): 19–21.
- 91 Kolly M, Pecoud A, Frei PC. Additives contained in drug formulations most frequently prescribed in Switzerland. *Annals of Allergy* 1989; **62**: 21–5.
- 92 Berlin CM, Jr, McCarver DG, Notterman DA, Ward RM, Weismann DN, Wilson GS, Wilson JT. 'Inactive' ingredients in pharmaceutical products: Update. *Pediatrics* 1997; **99**: 268–78.
- 93 Harvey SC. Antiseptics and disinfectants; fungicides, ectoparasiticides. *The Pharmacological Basis of Therapeutics*, 6th edn. New York: Macmillan, 1975: 1001–2.
- 94 Durieux ME, Neitgen GW. Synergistic inhibition of muscarinic signaling by ketamine stereoisomers and the preservative benzethonium chloride. *Anesthesiology* 1997; **86**: 1326–33.

- 95 Nordt SP. Chlorobutanol toxicity. *Annals of Pharmacotherapy* 1996; **30**: 1179–80.
- 96 DeChristoforo R, Corden BJ, Hood JC, Narang PK, Magrath IT. High-dose morphine infusion complicated by chlorobutanol-induced somnolence. *Annals of Internal Medicine* 1983; **98**: 335–6.
- 97 Fischer M. Effects of chlorobutanol on primary and secondary endings of isolated cat muscle spindles. *Brain Research* 2000; **854**: 106–21.
- 98 Simon RA. Adverse reactions to drug additives. *Journal of Allergy and Clinical Immunology* 1984; **74**: 623–30.
- 99 Routledge EJ, Parker J, Odum J, Ashby J, Sumpter JP. Some alkyl hydroxy benzoate preservatives (parabens) are estrogenic. *Toxicology and Applied Pharmacology* 1998; **153**: 12–9.
- 100 Pederson KL, Pederson SN, Christiansen LB, Korsgaard B, Bjerregaard P. The preservatives ethyl-, propyl- and butylparaben are oestrogenic in an in vivo fish assay. *Pharmacology and Toxicology* 2000; **86**: 110–13.
- 101 Harvey BH, Carstens ME, Taljaard JJ. Central effects of the preservative, methylparaben. In vivo activation of a cAMP-specific phosphodiesterase and reduction of cortical cAMP. *Biochemical Pharmacology* 1992; **44**: 1053–7.
- 102 Gall H, Kaufmann R, Kalveram CM. Adverse reactions to local anesthetics: Analysis of 197 cases. *Journal of Allergy and Clinical Immunology* 1996; **97**: 933–7.
- 103 Kajimoto Y, Rosenberg ME, Kytta J, Randell T, Tuominen M, Reunala T, Rosenberg PH. Anaphylactoid skin reactions after intravenous regional anaesthesia using 0.5% prilocaine with or without preservative – a double-blind study. *Acta Anaesthesiologica Scandinavica* 1995; **39**: 782–4.
- 104 Hamilton JT, Zhou Y, Gelb AW. Paraben preservatives but not succinylcholine are cerebral vasodilators in vitro. *Anesthesiology* 1990; **73**: 1252–7.
- 105 Brandt L, Andersson KE, Hindfelt B, Ljunggren B, Pickard JD. Are the vascular effects of naloxone attributable to the preservatives methyl- and propylparaben? *Journal of Cerebral Blood Flow and Metabolism* 1983; **3**: 395–8.
- 106 Gelb AW, Gignac E, Manninen PH, Farrar JK, Lee DH. Methylparaben and propylparaben do not alter cerebral blood flow in humans. *Canadian Journal of Anaesthesia* 1992; **39**: 691–4.
- 107 Pompy L, Karlin A, Capuano CM, Cottrell JE, Hartung J. Paraben preservatives do not increase intracranial pressure in cats. *Anesthesiology* 1991; **75**: 669–72.
- 108 Schnuch A, Geier J, Uter W, Frosch PJ. Patch testing with preservatives, antimicrobials and industrial biocides. Results from a multicentre study. *British Journal of Dermatology* 1998; **138**: 467–76.
- 109 Mowad CM. Allergic contact dermatitis caused by parabens: 2 case reports and a review. *American Journal of Contact Dermatitis* 2000; **11**: 53–6.
- 110 Simon RA. Adverse reactions to food additives. *New England & Regional Allergy Proceedings* 1986; **7**: 533–42.
- 111 Gould GW. Preservation past, present and future. *British Medical Bulletin* 2000; **56**: 84–96.
- 112 Sjoberg M, Karlsson PA, Nordberg C, et al. Neuropathologic findings after long-term intrathecal infusion of morphine and bupivacaine for pain treatment in cancer patients. *Anesthesiology* 1992; **76**: 173–86.
- 113 Hodgson PS, Neal JM, Pollock JE, Liu SS. The neurotoxicity of drugs given intrathecally (spinal). *Anesthesia and Analgesia* 1999; **88**: 797–809.
- 114 Hetherington NJ, Dooley MJ. Potential for patient harm from intrathecal administration of preserved solutions. *Medical Journal of Australia* 2000; **173**: 141–3.
- 115 Malinovsky JM, Lepage JY, Cozian A, Mussine JM, Pinaud M, Souron R. Is ketamine or its preservative responsible for neurotoxicity in the rabbit? *Anesthesiology* 1993; **78**: 109–15.
- 116 Gissen AJ, Datta S, Lambert D. The chloroprocaine controversy. II Is chloroprocaine neurotoxic? *Regional Anesthesia* 1984; **9**: 135–45.
- 117 Eisenach JC, Hood DD, Curry R. Phase I human safety assessment of intrathecal neostigmine containing methyl- and propylparabens. *Anesthesia and Analgesia* 1997; **85**: 842–6.
- 118 Hood DD, Eisenach JC, Tuttle R. Phase I safety assessment of intrathecal neostigmine in humans. *Anesthesiology* 1995; **82**: 331–43.
- 119 Adams HJ, Matri AR, Charron D. Morphological effects of subarachnoid methylparaben on rabbit spinal cord. *Pharmacology Research Communications* 1977; **9**: 547–51.
- 120 Mizuno K, Ogawa S, Itoh S. Suppressive effect of methylparaben on the evoked compound action potentials in excised rabbit cervical vagus nerve. *Masui* 1994; **43**: 1008–14.
- 121 Gurun MS, Leinbach R, Moore L, Lee C-S, Owen MD, Eisenach JC. Studies of the safety of glucose and paraben-containing neostigmine for intrathecal administration. *Anesthesia and Analgesia* 1997; **85**: 317–23.
- 122 Wang BC, Li D, Hiller JM, et al. Lumbar subarachnoid ethylenediaminetetraacetate induces hindlimb tetanic contractions in rats: prevention by CaCl₂ pretreatment – observation of spinal nerve root degeneration. *Anesthesia and Analgesia* 1992; **75**: 895–9.
- 123 Wills MH, Johns RA, Stone DJ, Moscicki JC, Difazio CA. Vascular effects of 2-chloroprocaine and sodium metabisulfite on isolated rat aortic rings. *Regional Anesthesia* 2000; **14**: 271–3.
- 124 Ravindran RS, Bond VK, Tasch MD, Gupta CD, Luerssen TG. Prolonged neural blockade following regional analgesia with 2-chloroprocaine. *Anesthesia and Analgesia* 1980; **59**: 447–51.
- 125 Hahn AF, Feasby TE, Gilbert JJ. Paraparesis following intrathecal chemotherapy. *Neurology* 1983; **33**: 1032–8.
- 126 Saiki JH, Thompson S, Smith F, Atkinson R. Paraplegia following intrathecal chemotherapy. *Cancer* 1972; **29**: 370–4.
- 127 Bagshawe KD, Magrath IT, Golding PR. Intrathecal methotrexate. *Lancet* 1969; **2**: 1258.

- 128 Gagliano RG, Costanzi JJ. Paraplegia following intrathecal methotrexate: report of a case and review of the literature. *Cancer* 1976; **37**: 1663–8.
- 129 Cousins MJ, Mather LE. Intrathecal and epidural administration of opiates. *Anesthesiology* 1984; **61**: 276–310.
- 130 Bromage PR, Camporesi E, Durant Pacielson CH. Rostral spread of epidural morphine. *Anesthesiology* 1982; **56**: 431–4.
- 131 Aldrete JA, Giles SL. Is the cost of preservative-free opioids for intraspinal use justified? *Journal of Pain and Symptom Management* 1997; **14**: 326–7.
- 132 Cherry DA, Gourlay GKCT Contrast evidence of injectate encapsulation after long-term epidural encapsulation. *Pain* 1992; **49**: 369–71.
- 133 Aldrete JA. Epidural fibrosis after permanent catheter insertion and infusion. *Journal of Pain Symptom Management* 1995; **10**: 624–31.
- 134 King CC, Hart LL. Epidural administration of methylprednisolone acetate preserved with benzyl alcohol. *Annals of Pharmacotherapy* 1994; **28**: 59–60.
- 135 Abram SE, O'Connor TC. Complications associated with epidural steroid injections. *Regional Anaesthesia* 1996; **21**: 149–62.
- 136 Bogduk N. Epidural steroids. *Spine* 1995; **20**: 845–8.
- 137 Johnson A, Ryan MD, Roche J. Depo-medrol and myelographic arachnoiditis. *Medical Journal of Australia* 1991; **155**: 18–20.
- 138 Nelson DA. Dangers from intraspinal steroid injections. *Archives of Neurology* 1990; **47**: 255.
- 139 Sekel R. Depo-medrol revisited. *Medical Journal of Australia* 1984; **140**: 688.
- 140 Spaccarelli K. Lumbar and caudal epidural corticosteroid injections. *Mayo Clinic Proceedings* 1996; **71**: 169–78.
- 141 Cicala RS, Turner R, Moran E, Henley R, Wong R, Evans J. Methylprednisolone acetate does not cause inflammatory changes in the epidural space. *Anesthesiology* 1990; **72**: 556–8.
- 142 Benzon HT, Gissen AJ, Strichtartz GR, Avran MJ, Covino BG. The effect of polyethylene glycol on mammalian nerve impulses. *Anesthesia and Analgesia* 1987; **66**: 553–9.
- 143 Delaney TJ, Rowlingson JC, Carron H, Butler A. Epidural steroid effects on nerves and meninges. *Anesthesia and Analgesia* 1980; **59**: 610–4.
- 144 Strong WE, Wesley R, Winne AP. Epidural steroids are safe and effective when given appropriately. *Archives of Neurology* 1991; **48**: 1012–13.
- 145 Pollock I, Young E, Stoneham M, Slater N, Wilkinson JD, Warner JO. Survey of colourings and preservatives in drugs. *British Medical Journal* 1989; **299**: 649–51.
- 146 Anonymous. Solu-Cortef and Solu-Medrol labels should indicate their preservative contents. *Hospital Pharmacology* 1993; **28**: 709.
- 147 Cooper MS. Preservative efficacy: compendial and regulatory issues. *Journal of Parenteral Science and Technology* 1989; **43**: 187–90.
- 148 Robinson JR. The need for new preservatives. *Journal of Parenteral Science and Technology* 1987; **41**: 143.